

Plotting experimental data using `pgfplots`

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Abstract

Creating plots in \TeX is made easy by the `pgfplots` package, but getting the best presentation of experimental results still requires some thought. In this article, the basics of `pgfplots` are reviewed before looking at how to adjust the standard settings to give both good looking and scientifically precise plots.

1 Introduction

Presenting experimental data clearly and consistently is a crucial part of publishing scientific results. A key part of this is the careful preparation of plots, graphs and so forth. Good quality plots often make results clearer and more accessible than large tables of numbers. A number of tools specialise in producing plots for scientific users, both commercial (such as `ORIGIN` and `SIGMAPLOT`) and open source (for example `QTIPLOT` and `SCIDAVIS`). The output of these programs is impressive, but \TeX users may find that it lacks the ‘polish’ that \TeX can provide.

There are a few approaches to producing plots directly within a \TeX document, but perhaps the easiest method to use the `pgfplots` package (Feuersänger, 2010). This is an extension of the very popular `pgf` graphics system (Tantau, 2008), which as many readers will know works equally well with the traditional DVI-based work flow and the increasingly popular direct production of PDF output. `pgfplots` also works with plain \TeX , \LaTeX and `ConTeXt`, meaning that it is an accessible route for almost all \TeX users.

As with any tool, getting the best results with `pgfplots` does require a bit of understanding. In this article, I’m going to look at getting good results for plots of experimental data. This includes things like worrying about units and how to get the graphics out of \TeX for publishers who require it.

In the examples, I am going to use real experimental data from the research group I work in,¹ and explain how I’ve presented this for publication. The aim is to show `pgfplots` in use ‘in the wild’, rather than the usual approach of somewhat contrived sets of data. The examples do not aim to be a complete survey of the power of `pgfplots`: its manual is excellent and covers all of the options in detail.

2 The basics

There are some basics that need to be in place before plots can be created. First, the code for `pgfplots`

¹ My supervisor is Professor Christopher Pickett:
<http://www.uea.ac.uk/che/pickettc>

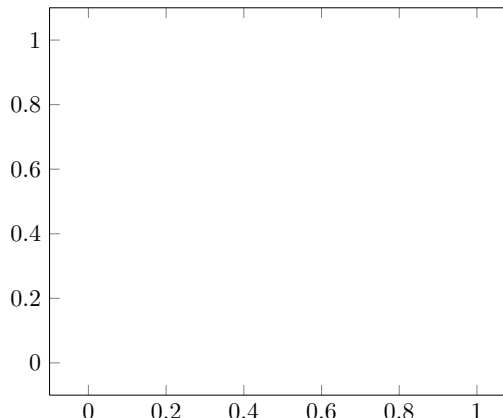


Figure 1: An empty set of axes

needs to be loaded. This is designed to work equally well with \TeX , \LaTeX and `ConTeXt`, and of course the loading mechanism depends on the format:

```
\input pgfplots.tex % Plain TeX
\usepackage{pgfplots} % LaTeX
\usemodule[pgfplots] % ConTeXt
```

Plots are created inside an ‘`axis`’ environment, which itself needs to be inside the `pgf` environment ‘`tikzpicture`’. The differences between the three formats again mean that there are again some variations. So, for plain \TeX :

```
\tikzpicture
  \axis
    % Plot code
  \endaxis
\endtikzpicture
```

while for \LaTeX :

```
\begin{tikzpicture}
  \begin{axis}
    % Plot code
  \end{axis}
\end{tikzpicture}
```

and for `ConTeXt`:

```
\starttikzpicture
  \startaxis
    % Plot code
  \stopaxis
\stoptikzpicture
```

In the examples in this article I’ll leave out the wrapper and concentrate just on the plot code, which is the same for all three formats.

If the wrapper is given with no content at all then `pgfplots` will fill in some standard values. This gives an empty plot (Figure 1). To actually have something appear, data has to be added to the axes. This is done using one or more `\addplot` instructions, which have a flexible syntax which can be applied to a wide range of cases.

Table 1: Rates of a chemical reaction [Data taken from Jablonskytė, Wright, and C. J. Pickett (2010)].

Concentration / mmol dm ⁻³	Rate / s ⁻¹
338.1	266.45
169.1	143.43
84.5	64.80
42.3	34.19
21.1	9.47

Before starting to produce some real plots, there is one minor thing to set up. The most recent version of `pgfplots` (1.3) makes a number of improvements to the standard settings for the package, but only if told to. For new plots, it makes sense to use these improved abilities using

```
\pgfplotsset{compat = newest}
```

in the source. In this article, I've included this line in the preamble for the source (which is in L^AT_EX).

3 Small data sets

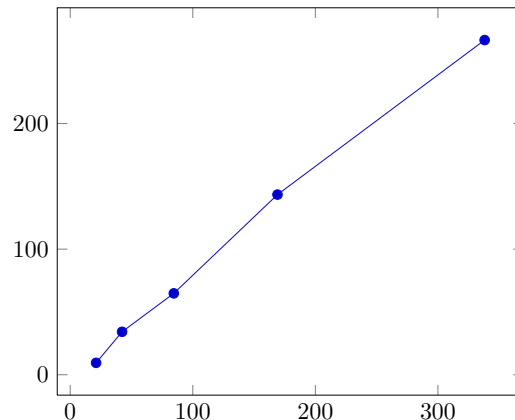
Small sets of data, for example values calculated by hand, can be plotted by including the data directly in the T_EX source. As an example set of data, I am going to use the rate of a chemical reaction, recently reported by the research group I work in (Table 1). As shown, the rate of a chemical reaction depends on the concentration of one of the 'ingredients', which as a table is suggestive but not really immediately accessible.

This can be plotted rapidly using the `\addplot` macro along with the `coordinates` keyword, with the input reading

```
\addplot coordinates {
  ( 338.1, 266.45 )
  ( 169.1, 143.43 )
  ( 84.5, 64.80 )
  ( 42.3, 34.19 )
  ( 21.1, 9.47 )
};
```

(placed inside the `axis` environment, as explained above). This shows a number of features of the `pgfplots` input syntax. First, after the primary macro (`\addplot`) a keyword is used to direct the behaviour of the code. Second, there is no need to worry about white space, which makes it easier to read the input. Third, as with other `pgf` commands, the entire `\addplot` input is terminated by a semi-colon. This will allow multiple plots to use the same axes, as will be demonstrated later.

Using the input above also reveals why it is necessary to think about plots, even with a powerful

**Figure 2:** A raw plot based on the data in Table 1.

system like `pgfplots`. Using the simple input I've given results in Figure 2. There are a number of issues with this initial plot. Most obviously, the data points should not be plotted in a 'join the dots' fashion. Much better would be independent points along with a best fit line showing the trend. The plot is also in colour, which for a simple plot like this one is not really necessary. Publishers prefer black and white unless colour is adding real information.

As with other parts of the `pgf` system, `pgfkeys` uses key-value arguments to alter settings. Here, the options apply to a particular plot, and so are given as an optional argument to the `\addplot` macro in square brackets (irrespective of the format in use). So the appearance of the plot can be altered by adding an optional argument and suitable settings to the `\addplot` macro.

```
\addplot [
  color = black,
  fill = black,
  mark = *, % A filled circle
  only marks
]
```

These keys all have self-explanatory names: the `pgfplots` manual of course gives full details. It is possible to use 'black' for the combination of `color = black` and `fill = black`; here, I will stick to the longer version including the key names, as it makes the meaning a little clearer.

Adding a best fit line means a second `\addplot` is needed. T_EX is not the best way to do general mathematics, and so the fitting was done using a spreadsheet application. The easiest way to add a line is to specify the two ends and show only the line:

```
\addplot [
  color = black,
  mark = none
]
```

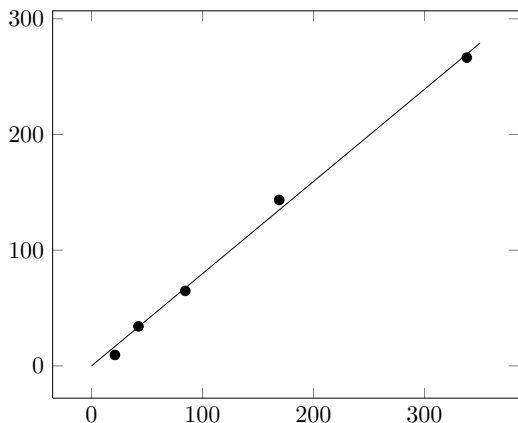


Figure 3: Second version of a plot based on the data in Table 1.

```
coordinates {
  ( 0, 0 )
  ( 350, 279 )
};
```

Making these first set of changes gives Figure 3, which already looks more professional.

There is still more to think about before the plot is finished. The axes do not start from 0, but instead from a bit below zero: hardly very clear, and certainly not needed here. Much more importantly, the axes are not labelled and there are no units. These are both problems about the entire plot, not just one data set. So in this case the optional argument to the `axis` environment comes into play. This follows the start of the environment in square brackets (recall that the start of the environment is format-dependent, as shown above). Once again the various option names are pretty self-explanatory:

```
[
  xlabel = Concentration\,/\,mmol\,dm$^{-3}$,
  xmax   = 400,
  xmin   = 0,
  ylabel = Rate\,/\,s$^{-1}$,
  ymax   = 300,
  ymin   = 0
]
```

The result of these adjustments is Figure 4. The contents of the `axis` environment is now:

```
[
  xlabel = Concentration\,/\,mmol\,dm$^{-3}$,
  xmax   = 400,
  xmin   = 0,
  ylabel = Rate\,/\,s$^{-1}$,
  ymax   = 300,
  ymin   = 0
]
```

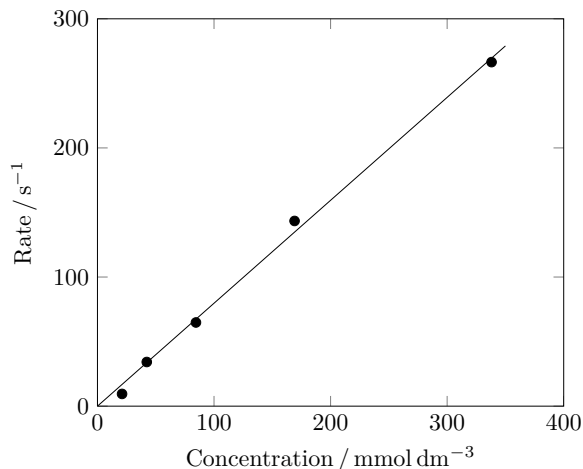


Figure 4: Final version of a plot based on the data in Table 1.

```
\addplot[
  color = black,
  fill  = black,
  mark  = *,
  only marks
] coordinates {
  ( 338.1, 266.45 )
  ( 169.1, 143.43 )
  ( 84.5, 64.80 )
  ( 42.3, 34.19 )
  ( 21.1, 9.47 )
};
\addplot[
  color = black,
  mark  = none
] coordinates {
  ( 0, 0 )
  ( 350, 279 )
};
```

The plot now looks good, but there are a few other points to note before moving on to more complex challenges. In a real publication, the caption of the figure should give any necessary experimental details about the data presented. Also notice that both axes of the plot are in simple whole numbers. For this plot (and Table 1), the original concentrations were in mol dm^{-3} , and were multiplied by 1000 to make them whole numbers. Usually this approach makes the final result much easier to read (even if it requires more effort initially). `pgfplots` can do simple mathematics, but I tend to favour doing the calculations first in an external tool and stick to using `TEX` for its strength: typesetting.

The ideas used for the preceding plot can readily be applied to presenting several sets of data on the same axes. `pgfplots` allows the user to pick a number

of markers to differentiate the plots, and to include legends and so forth. However, it rapidly becomes unwieldy to have the raw data in the \TeX source when there are a number of curves to plot. Luckily, `pgfplots` has an alternative `\addplot` syntax that help out.

4 Large sets of data

As the number of data points to plot grows, adding the information directly to the source rapidly becomes laborious and error-prone. Almost always, the data will be available in a structured format, and so reading an external file becomes a much more efficient way to proceed. `pgfplots` can read data tables from, for example, tab-separated text files, which can conveniently be written by standard data handling software. There are two approaches to reading such data: loading the entire table into a macro which can then be used for plotting, or reading the data as part of the `\addplot` line. The most appropriate method depends on the context: both methods will be illustrated here.

4.1 One set of axes, several plots

When several related sets of data need to be presented on one set of axes, it rapidly becomes easier to use external data files even if each plot has only a small number of points.

Here, I will illustrate the methods using the change in amount of four chemicals over time. The data here are available as a single text file, and there are not too many time points. It therefore makes most sense to load all of the data into a macro, and to use it to create each plot in turn. With the numbers saved in a file called `data-set-two.txt`, the reading instruction is

```
\pgfplotstableread{data-set-two.txt}
  \datatable
```

Here, the name of the storage macro (`\datatable`) is arbitrary: in a complex document it would probably be more descriptive.

In the table here each column has a header for ease of reference, with the first few rows reading

Time	a	b	c	d
0	49	7	41	1.3
67	55	9	33	1.6
134	61	10	26	1.9
200	65	12	20	1.9
...				

The columns can be referred to either by their header name (`'y = <header name>'`) or by their position in the table (`'y index = <header index>'`), with column 'a' here being the y column with index 1, 'b' with index 2, and so on. It's also possible to include

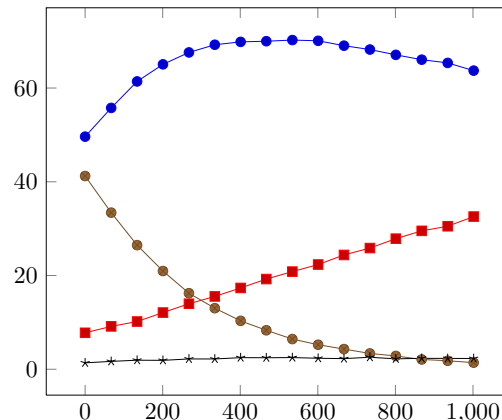


Figure 5: Raw plot using a pre-loaded data table.

comment lines in the data file, which can be used for things like the units or reference numbers for the raw data.

The plot can then be created using the 'table' keyword after the `\addplot` macro:

```
\addplot table[y = a] from \datatable ;
\addplot table[y = b] from \datatable ;
\addplot table[y = c] from \datatable ;
\addplot table[y = d] from \datatable ;
```

Here, the second keyword 'from' indicates that a macro will be used to supply the data to plot. With no formatting changes the result is Figure 5. As with the first data set, it is better not to 'join the dots'. As this applies to the entire plot, **only marks** can be given in the optional argument to the start of the `axis` environment, rather than repeating the same instruction for each `\addplot` macro. There also needs to be a legend to tell the reader which curve is which. Using the `\addlegendentry` macro after each `\addplot` line will automatically gather the necessary information (symbol type and colour), and will result in a legend being added to the plot:

```
\addplot table[y = a] from \datatable ;
\addlegendentry{Compound \textbf{a}} ;
\addplot table[y = b] from \datatable ;
\addlegendentry{Compound \textbf{b}} ;
...
```

The ideas about labelling axes and setting an appropriate scale which were necessary for the first plot also apply here. Making these adjustments leads to Figure 6.

Once again, colour is not really necessary here if care is taken with the rest of the plot. The visual difference between the different markers should be enough to show which curve is which. When preparing the real diagram for publication, my supervisor asked for all of the symbols to be circles, filled

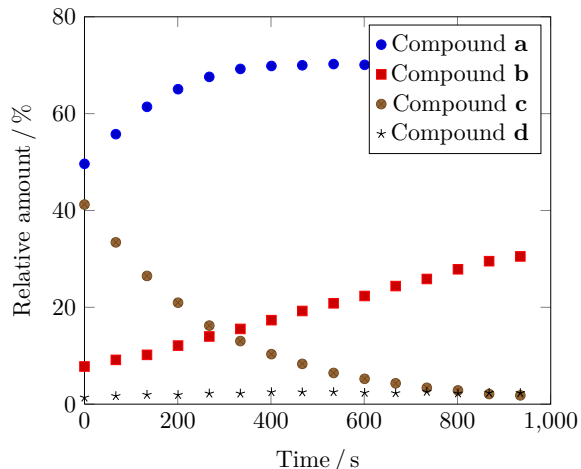


Figure 6: Second version of a plot from a pre-loaded data table.

in different ways. That needed a bit of help from `comp.text.tex` to get half-filled circles, which can be achieved by creating a special marker using some lower level `pgf` code:

```
\pgfdeclareplotmark{halfcircle}{%
  \begin{pgfscope}
    \pgfsetfillcolor{white}%
    \pgfpathcircle{\pgfpoint{0pt}{0pt}}
      {\pgfplotmarksizes}
    \pgfusepathqfillstroke
  \end{pgfscope}%
  \pgfpathmoveto
    {\pgfpoint{\pgfplotmarksizes}{0pt}}
  \pgfpatharc{0}{180}{\pgfplotmarksizes}
  \pgfpathclose
  \pgfusepathqfill
}
```

(don't worry too much about this; for myself, I just accept that it works!). Including the above code in the source means that `halfcircle` can be used as a value for the `mark` key:

```
\addplot[mark = halfcircle, ...
```

The filled portion can be moved 'around' the circle by rotating the mark

```
\addplot[
  mark = halfcircle,
  mark options = {rotate = 90},
```

There are a couple of other issues with Figure 6. Most obviously, the legend is covering some of the data points, while there is a handy space right in the middle. This can be fixed by asking `pgfplots` to move the entire legend box using the `legend style` key, which is used in the optional argument to the axis environment. It takes a bit of experimentation to get the correct position; in this case

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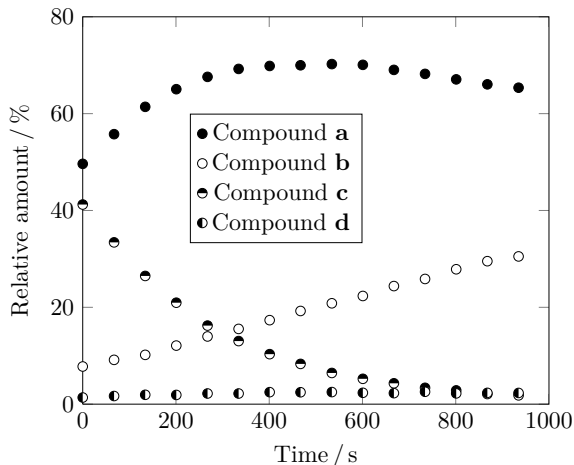


Figure 7: Final version of a plot from a pre-loaded data table.

```
[
  legend style = { at = {(0.6,0.75)}}
]
```

seems to be about right.

The x axis label for 1000 includes a comma as a digit separator: that is not usual in publications in English. So there is a slight modification to make to the digit formatting routine: this is a general `pgf` setting:

```
\pgfkeys{
  /pgf/number format/
  set thousands separator =
}
```

Making these changes, and adjusting a few minor settings to get the colours correct leads to the final version of the plot (Figure 7). Here is the code:

```
[
  legend style = { at = {(0.6,0.75)}},
  only marks,
  xlabel = Time\,/ \,s,
  xmax = 1000,
  xmin = 0,
  ylabel = Relative amount\,/ \,%,
  ymax = 80,
  ymin = 0
]
\addplot[
  color = black,
  mark = *
] table[y = a] from \datatable ;
\addlegendentry{Compound \textbf{a}} ;
\addplot[
  color = black,
  fill = white,
  mark = *
] table[y = b] from \datatable ;
```

```

\addlegendentry{Compound \textbf{b}} ;
\addplot[
  color      = black,
  mark       = halfcircle
] table[y = c] from \datatable ;
\addlegendentry{Compound \textbf{c}} ;
\addplot[
  color      = black,
  mark       = halfcircle,
  mark options = {rotate = 90}
] table[y = d] from \datatable ;
\addlegendentry{Compound \textbf{d}} ;

```

4.2 An experimental spectrum

One common technique in scientific research is recording *spectra*: how a sample absorbs light, microwaves, radio waves, *etc.* Often, these are published by simply taking the raw print out from the control program and pasting it into the article, which does not make for a good appearance. So replotting with `pgfplots` is a good idea.

Spectra tend to involve a lot of numbers, and so using an external table is once again a good idea. As these are large files that will only be used once, loading to a macro is not efficient. Instead, the data table can be loaded as part of the `\addplot` line, with the syntax

```
\addplot table {data-set-three.txt};
```

The first version of the plot in this case (Figure 8) already includes a number of the refinements discussed in the first two examples. The example shows data from a technique called ‘nuclear magnetic resonance’, in which radio waves are absorbed by a sample. In the plot, the x axis is related to the frequency of the radio waves, while the y axis shows how much is absorbed.

The plot looks generally good, but there are some adjustments required. First, the y scale is essentially arbitrary, and so is usually not given any values at all. This can be achieved by setting the `yticklabels` option to an empty value in the optional argument to the `axis` environment:

```
[yticklabels = ]
```

Second, for various historical reasons it is normal to plot the x axis backward (running high to low). The latest version of `pgfplots` can do this automatically using the `x dir` option.

```

\addplot[
  x dir = reverse,
  ...

```

These changes give an improved version of the plot (Figure 9).

The final thing this plot needs is some ‘peak labels’: markers showing the exact value at the top

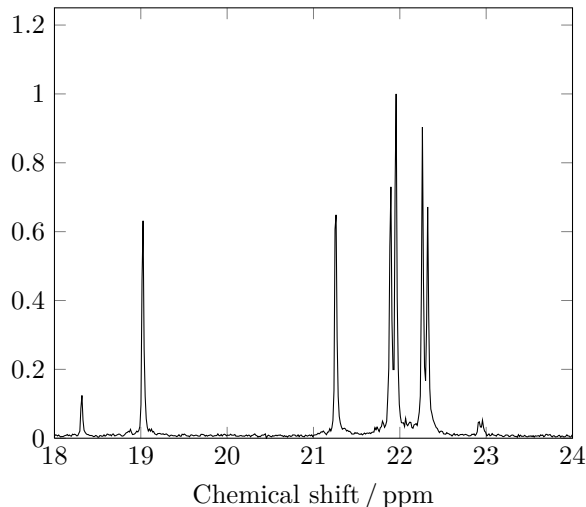


Figure 8: First version of a single spectrum.

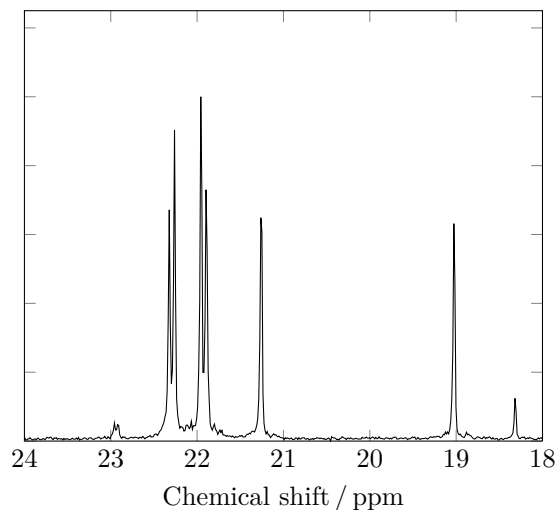


Figure 9: Second version of a single spectrum.

of each peak. For this, the `pgf \node` macro can be used. A `\node` can be used to put material anywhere on a plot, and each node takes a range of options to control its appearance. In this case, the node itself is going to be completely empty, and a `pin` will be used to point to the node. A `pin` generates a short line from the node to some associated text, which in this case I want to rotate by 90° to pack the labels in. The syntax ends up a little bit complicated, as various options need to be correct, for example:

```

\node[
  coordinate,
  pin = {[rotate=90]right:22.26}
] at (axis cs:22.26,1.1) { };

```

The `right` keyword here can be replaced by an angle, which can then be used to be slightly off a peak's position, for example

```
\node[
  coordinate,
  pin = {[rotate=90]5:22.32}
] at (axis cs:22.32,1.1) { };
```

Once again, putting everything together leads to the completed plot (Figure 10).

```
[
  x dir      = reverse,
  xlabel     = Chemical shift\,/,\,ppm,
  xmin      = 18,
  xmax      = 24,
  ymax      = 1.75,
  ymin      = 0,
  yticklabels =
]
\addplot[
  color = black,
  mark = none
] table from {data-set-three.txt};
\node[
  coordinate,
  pin = {[rotate=90]5:22.32}
] at (axis cs:22.32,1.1) { };
\node[
  coordinate,
  pin = {[rotate=90]right:22.26}
] at (axis cs:22.26,1.1) { };
\node[
  coordinate,
  pin = {[rotate=90]right:21.96}
] at (axis cs:21.96,1.1) { };
\node[
  coordinate,
  pin = {[rotate=90]-5:21.90}
] at (axis cs:21.90,1.1) { };
\node[
  coordinate,
  pin = {[rotate=90]right:21.26}
] at (axis cs:21.26,1.1) { };
\node[
  coordinate,
  pin = {[rotate=90]right:19.03}
] at (axis cs:19.03,1.1) { };
\node[
  coordinate,
  pin = {[rotate=90]right:18.32}
] at (axis cs:18.32,1.1) { };
```

4.3 Changes over time

The previous example demonstrated how to plot a single spectrum from an external file. In a final example, I want to look at going beyond this to showing how experimental data changes over time. This means plotting a series of spectra on the same

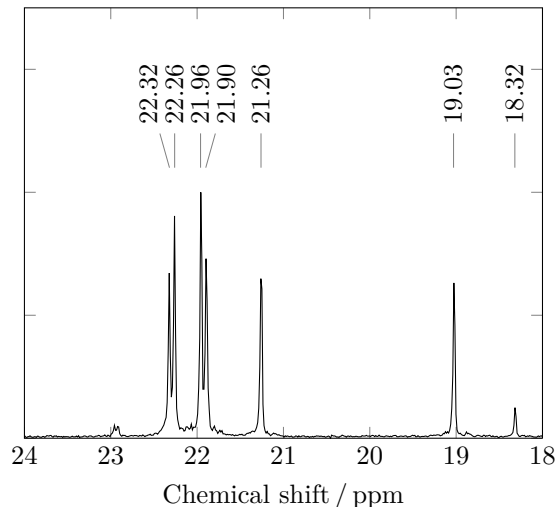


Figure 10: Final version of a single spectrum.

axes, and finding a good way to show which way time is running. Some people choose to use a ‘three-dimensional’ plot for this scenario, and `pgfplots` includes the ability to generate this type of output. However, experience suggests to me that there is more value in a well-constructed two-dimensional plot than a three-dimensional representation of the same data. The challenge is therefore to find the best way to represent the results on paper.

In this example, the raw data is how a sample absorbs infra-red light (heat). There is a reaction taking place, and so the absorption will change over time. The initial results simply show a series of peaks, a bit like Figure 10. The changes are quite subtle compared to the overall scale, so the first stage in creating a plot is not `TEX` related. Using a spreadsheet it’s possible to find how the signal changes relative to the one at the start of the experiment: this gives a ‘difference spectrum’ for each time. That can then be saved as a text file which can be used as the input to `pgfplots`.

In this case, a single file contains all of the data for the plot. To get each `\addplot` line to use a single time, the optional argument to the `table` keyword is used, for example:

```
\addplot[mark = none] table[y index = 1]
  {data-set-four.txt};
\addplot[mark = none] table[y index = 2]
  {data-set-four.txt};
...
```

As there are many almost identical lines, the `pgf` `\foreach` macro can be used to vary only the `y` index used

```
\foreach \yindex in {1,2,...,20}
  \addplot[mark = none]
```

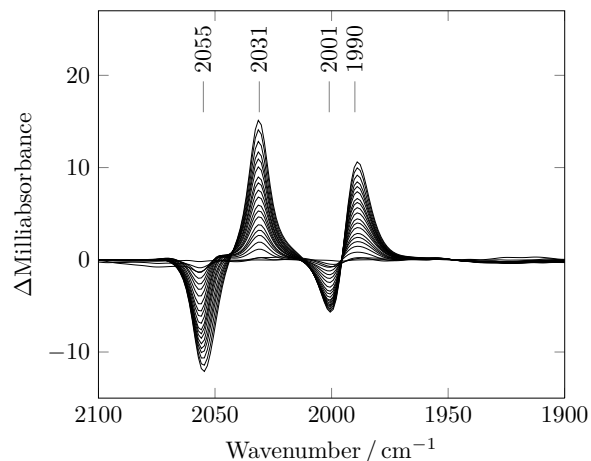


Figure 11: First plot of a change over time.

```
table[y index = \yindex]
{data-set-four.txt};
```

A bit of experimentation showed that around 20 lines gave a good appearance for the plot (the full data set has nearly 200 time points!). The `\foreach` syntax makes it easy to show an evenly-spaced subset of the available points, by setting the gap between selected columns to be larger:

```
\foreach \yindex in {1,10,...,189}
\addplot[mark = none]
table[y index = \yindex]
{data-set-four.txt};
```

This uses every 9th column for the plot, and in the example results in 19 separate curves (Figure 11).

There is one obvious problem with the plot: which way is time running? Here, careful use of colour can be used in a way which really does add to the information imparted by the plot. To do this, a ‘plot cycle’ is needed to specify the colour used for each plot. There are some built in to `pgfplots`, but one is also easy to construct:

```
\pgfplotscreateplotcyclelist
{blue to red}{%
color = red!0!blue%%
color = red!5!blue%%
color = red!10!blue%%
color = red!15!blue%%
color = red!20!blue%%
color = red!25!blue%%
color = red!30!blue%%
color = red!35!blue%%
color = red!40!blue%%
color = red!45!blue%%
color = red!50!blue%%
color = red!55!blue%%
color = red!60!blue%%
color = red!65!blue%%
```

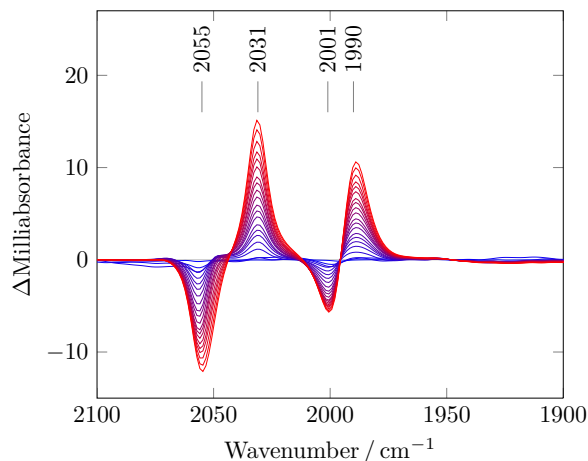


Figure 12: Second plot of a change over time (time runs blue to red).

```
color = red!70!blue%%
color = red!75!blue%%
color = red!80!blue%%
color = red!85!blue%%
color = red!90!blue%%
color = red!95!blue%%
color = red!100!blue%%
}
```

It’s important to note that the end of lines here must end in comments. With that cycle available, the option `cycle list name` can be used for the axes: this will mean that the first plot will be pure blue, the second 95% blue and 5% red, and so on.

The other minor adjustment to make is to include a line at for $y = 0$. This can be done by including an extra ‘tick’:

```
[
extra y ticks      = 0,
extra y tick labels = ,
extra y tick style =
{ grid = major }
]
```

(The ‘`extra y tick labels = ,`’ line makes sure that the label for 0 is not printed twice, as this gives a slightly ‘bold’ appearance.)

These two adjustments lead to Figure 12:

```
[
cycle list name    = blue to red,
extra y ticks      = 0,
extra y tick labels = ,
extra y tick style = { grid = major },
x dir              = reverse,
xlabel             = Wavenumber\,/ \, cm$^{-1}$,
xmin              = 1900,
xmax              = 2100,
ylabel            = $ \Delta $Milliabsorbance,
```



```

ymax          = 27,
ymin          = -15
]
\foreach \yindex in {1,10,...,189}
  \addplot table[y index = \yindex]
    {data-set-four.txt};
\node[
  coordinate,
  pin = {[rotate=90]right:1990}
] at (axis cs:1990,16) { };
\node[
  coordinate,
  pin = {[rotate=90]right:2001}
] at (axis cs:2001,16) { };
\node[
  coordinate,
  pin = {[rotate=90]right:2031}
] at (axis cs:2031,16) { };
\node[
  coordinate,
  pin = {[rotate=90]right:2055}
] at (axis cs:2055,16) { };

```

As with the other plots, it's important to include details about the experiment somewhere close to the figure: usually the caption is a good place for this. In the published article, I included details about what I used as time zero, the time range and concentrations for the experiment in the caption.

5 Exporting plots from T_EX

Processing a large number of plots in T_EX can lead to the program running out of memory. The ability to set up plots as separate files which can then be included in the main document as graphics is therefore important. At the same time, some publishers require all graphics to be available as stand-alone files, which again means finding a way to export plots from T_EX.

pgfplots includes methods for carrying out this process in an automated fashion. The latest release of pgfplots makes the process much easier than was previously the case, and includes full instructions on how to proceed. When using pdfT_EX the lines

```

\usetikzlibrary{pgfplots.external}
\tikzexternalize{<filename>}

```

will instruct pgfplots to make each plot into a separate PDF file. To create PostScript files the additional command:

```

\tikzset{external/system call =
  {latex -shell-escape -halt-on-error
  -interaction=batchmode -jobname "\image"
  "\texsource"; dvips -o "\image".ps
  "\image".dvi}}

```

is needed, and the main file should be typeset in DVI mode. In both cases \write18 (external command execution) needs to be enabled for the process to work correctly.

6 Conclusions

Producing scientific plots using pgfplots can produce high quality output with relatively little effort for the user. To do so, some thought about both the information to be reported and the appearance of the output is needed.

7 Acknowledgements

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The published versions of the figures used here originally appeared in Wright and Pickett (2009) and Jablonskytė, Wright, and C. J. Pickett (2010). They are reproduced by permission of The Royal Society of Chemistry.

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